- primary structure of human chromogranin A (secretory protein I) cDNA. J biol Chem 1988, 263, 11559-11563.
- Brodeur GM, Pritchard J, Berthold F, et al. Revisions of the international criteria for neuroblastoma diagnosis, staging and response to treatment. J Clin Oncol 1993, 11, 1466-1477.
- Kogner P, Björk O, Theodorsson E. Neuropeptide Y as a marker in pediatric neuroblastoma. *Pediat Path* 1990, 10, 207–216.
- Kogner P, Björk O, Theodorsson E. Neuropeptide Y in neuroblastoma: increased concentration in metastasis, release during surgery and characterization of plasma and tumor extracts. *Med Ped Oncol* 1993, 21, 317-322.
- Kogner P, Björk O, Theodorsson E. Plasma neuropeptide Y in healthy children; influence of age, anaesthesia, and the establishment of an age-adjusted reference interval. Acta Paediat 1994, 83, 423-427.
- Kogner P, Björk O, Dominici C, Hedborg F, Theodorsson E. Vasoactive intestinal peptide (VIP) and somatostatin in childhood ganglioneuromas and neuroblastomas. *Proc Am Soc Clin Oncol* 1992, 11, 371.
- 11. Iguchi H, Funakoshi A, Tateishi K, Ichinose Y, Hara N, Ohta M. Production of pancreastatin-like immunoreactivity, a presumed processing product of chromogranin A, in small cell lung carcinoma. Cancer J 1990, 3, 197-201.

- 12. Iguchi H, Bannai S, Takanashi N, Tsukada Y. Production of chromogranin A and B derived peptides in human small cell lung carcinoma cell lines. *Eur J Cancer* 1992, 28A, 1458–1462.
- 13. Hsiao RJ, Seeger RC, Yu AL, O'Connor DT. Chromogranin A in children with neuroblastoma. Serum concentration parallels disease stage and predicts survival. J Clin Invest 1990, 85, 1555-1559.
- 14. Cooper MJ, Hutchins GM, Cohen PS, Helman LJ, Mennie RJ, Israel MA. Human neuroblastoma tumor cell lines correspond to the arrested differentiation of chromaffin adrenal medullary neuroblasts. Cell Growth Different 1990, 1, 149-159.
- Hachitanda Y, Tsuneyoshi M, Enjoji M. Expression of pan-neuroendocrine proteins in 53 neuroblastic tumors. An immunohistochemical study with neuron-specific enolase, chromogranin, and synaptophysin. Arch Path Lab Med 1989, 113, 381-384.
- Molenaar WM, Baker DL, Pleasure D, Lee VMY, Trojanowski JQ. The neuroendocrine and neural profiles of neuroblastomas, ganglioneuroblastomas and ganglioneuromas. Am J Path 1990, 136, 375-382.

Acknowledgements—The present work was supported by the Swedish Child Cancer Fund, the Swedish Cancer Society and Research Funds of the Karolinska Institute. We wish to acknowledge the expert technical assistance of Ms Anita O'Flaherty.

European Journal of Cancer Vol. 31A, No. 4, pp. 560-564, 1995 Copyright © 1995 Elsevier Science Ltd Printed in Great Britain. All rights reserved 0959-8049/95 \$9.50+0.00



0959-8049(95)00062-3

Genetic Alterations Associated with Metastatic Dissemination and Chemoresistance in Neuroblastoma

J. Bénard

Knowledge about genetic alterations specific to the metastatic process and chemoresistance in neuroblastoma is progressing steadily. Low or no CD44 expression, increased NM23 expression and specific mutations of the 5' coding regions of NM23 are distinct features of aggressive, metastatic neuroblastoma. MYCN down-regulates Class I HLA antigen expression in many neuroblastoma cell lines and, in turn, may be regulated by a suppressor gene. The MYCN amplified human neuroblastoma cell line, IGR-N-91, established in vitro, metastasises in the nude mouse and has exhibited co-activation of MYCN and PGY1, resulting from direct activation of the oncoprotein on the PGY1 promoter. In this model, the MYCN product activates angiogenesis, the dissemination process and chemoresistance via specific genes (PGY1 and GST3). MYCN, like the BCL-2 and TP53 products, may also play a key role in apoptosis. The implication of these genes in the potential for metastasis and chemoresistance in neuroblastoma is discussed.

Key words: neuroblastoma, metastatic dissemination, genetic alterations, MYCN activation, chemoresistance, in vivo models

Eur J Cancer, Vol. 31A, No. 4, pp. 560-564, 1995

INTRODUCTION

THE GENETIC analysis of tumour tissues has offered considerable insights into the tumour heterogeneity of neuroblastoma. Indeed, cytogenetic and molecular biology studies, currently in progress, have identified recurrent genetic alterations which, when combined, appear to denote the existence of neuroblastoma subtypes [1, 2]. In the clinic, knowledge of such genetic anomalies has provided immediate applications for the treatment and

outcome of patients with localised forms and stage IV-S [1, 3, 4]. So far, however, there has been no impact on the management of clinically unfavourable forms (disseminated stage IV) of neuroblastoma at diagnosis, in children older than a year. It is possible that information on genetic alterations specific to metastatic cells could help to improve the prognosis and to tailor therapy to these aggressive neuroblastomas.

Cancer invasion and dissemination is characterised by a long

series of sequential, inter-related steps [5] consisting, at a physiopathological level, of angiogenesis, invasion, intravasion, transport, arrest, extravasation, and proliferation to metastatic sites. [6] Metastatic dissemination is generally regarded as a late event in tumour progression [7] which, at the molecular level, is the gradual accumulation of multiple genetic changes affecting cell growth and neoplastic behaviour. There is considerable evidence to support the notion that tumour progression may be due to the activation, mutation or loss of different genes which belong to three major functional groups [8], i.e., oncogenes, suppressor genes and modulator genes. As proposed by Klein and Klein [8], the modulators seem to influence the neoplastic behaviour of tumour cells during their dissemination and interaction with host tissues. These genes form a large heterogeneous group, and include the genes of the major histocompatibility complex, those expressed in the control of proteolytic and homing mechanisms, genes involved in cellular resistance to immune rejection as well as those activated during resistance to treatment.

This review will present and discuss (i) the genetic alterations which could be specifically associated with metastasis in neuroblastoma; (ii) experimental models used to detect these genetic abnormalities; and (iii) the role of genes involved in the response of neuroblastoma cells to chemotherapy.

INTRINSIC GENETIC AND MOLECULAR DETERMINANTS RELATED TO TUMOUR PROGRESSION AND THE METASTATIC PROCESS IN NEUROBLASTOMA

Acquired recurrent genetic alterations characterise primary neuroblastoma. Mostly, these include loss of heterozygosity (LOH) on the short arm of chromosome 1, at band 1p36-2 [9, 10], MYCN proto-oncogene amplification [11, 12], hyperdiploidy or near diploidy [3] and defects in the expression or function of the nerve growth factor receptor [13]. Hyperdiploidy, i.e., increased DNA content, is associated with early stages of the disease and with a favourable outcome in infants. LOH for chromosome 1, and MYCN amplification are more common in children older than 1 year of age with advanced stages of disease: these two latter genetic abnormalities appear to be related [14]. LOH for 1p36 may precede MYCN gene amplification. By combining the DNA content, 1p LOH and MYCN amplification, 3 distinct genetic subtypes of neuroblastoma can be defined. The first group has a hyperdiploid modal karyotype, the second group, a near-diploid mode karyotype and the third, a diploid mode, 1p deletions or LOH for 1p36 and MYCN amplification. These three groups correspond to the different levels of prognosis (good, intermediate and very poor, respectively) and typify three distinct subtypes of neuroblastoma [1]. Of great importance, no connection exists between the 3 groups and molecular proof of a stepwise transition from one neuroblastoma type to another is lacking. Regarding the third group, which corresponds to advanced aggressive neuroblastoma, it is noteworthy that 1p deletion and MYCN amplification have been found in the primary tumour prior to metastatic dissemination [2]. If amplification of the MYCN gene is correlated with increased metastatic potential [12], the mechanism enabling MYCN to increase neuroblastoma malignancy is poorly understood. However, evidence has recently been presented that MYCN disrupts protein kinase C-mediated signal transduction in neuroblastoma [15]. Other alterations of oncogenes and growth factors, such as nerve growth factor (NGF) receptor [13, 16] and HA-RAS [17] expression have recently been reported and are additional elements regarding disease characterisation but so far their association with a potential for metastasis has not been determined.

More extensive research has been conducted for the heterogeneous class of modulator genes, possibly involved in the neuroblastoma metastatic process, and results obtained invariably refer to MYCN expression. The human CD44 cell surface integral glycoprotein, involved in a variety of functions, including lymphocyte homing, extracellular cell matrix attachment, and tumour metastasis, and subjected to alternative splicing, has been described as overexpressed in metastatic colon cancer [18]. In neuroblastoma, low expression or the absence of CD44 in cell lines strongly suggested that, unlike that found in other tumours, repressed CD44 expression could be a marker of aggression [19]. Two recent studies using immunohistochemical [20] and molecular [21] approaches confirmed this seminal work. In human tumours, CD44 is highly expressed in its standard isoform in 100% of stage I-III, stage IV-S neuroblastomas and ganglioneuromas, but only in a subset of stage IV tumours [21]. In contrast, no CD44 expression was detected on MYCN amplified stage IV tumours, indicating a highly negative relationship between MYCN amplification and CD44 expression in neuroblastoma [21]. In a recent multivariate analysis, CD44 expression appeared to be a marker of good prognosis [20].

Reduced expression of the NM23-H gene, which encodes the nucleoside diphosphate kinase, is associated with a high potential for metastasis in some tumour types, but this expression is increased in aggressive neuroblastoma [22]. Recent studies on mutations in primary tumours at different stages showed that specific mutations in the NM23-H gene 5' coding regions, i.e., leu $48 \rightarrow val$ [23] and ser $120 \rightarrow Gly$ [24] were frequently and specifically present in advanced tumours, but not in any early stage tumours. Altogether, these data indicate that molecular alterations to NM23, other than reduced expression, can be associated with tumour aggressiveness, and imply that the mutation could be a feature of advanced neuroblastomas.

The expression of Class I HLA antigens, membrane proteins that play an critical role in the recognition of tumour cells by cytotoxic T cells and NK cells, is down-regulated in many neuroblastoma lines. Class I HLA genes appear to be governed by the MYCN gene [25, 26] which, in turn, could be regulated by a suppressor gene [27].

IN VITRO ESTABLISHED MYCN AMPLIFIED HUMAN NEUROBLASTOMA CELL LINES METASTASISE IN THE NUDE MOUSE

Many neuroblastoma cell lines have been established in vitro from tumour tissue or involved bone marrow [28], the most frequent site of metastases in patients. In our laboratory, two MYCN-amplified neuroblastoma lines were both derived in vitro from stage IV disease, the former from a human primary tumour, IGR-N-835 [29], the latter from bone marrow metastases, IGR-N-91 [30], from another patient. Both IGR-N-91 and IGR-N-835 lines, which generated large tumours after subcutaneous (s.c.) xenografting on to nude mice, were able to disseminate and induce macroscopic metastases in the kidneys, adrenals and in lymphatic tissue of the mouse. In addition, occult neuroblastoma cells were present in the blood, the bone marrow and in the myocardium of animals, and could be characterised through in vitro organ culture. Subculturing of

J. Bénard

these neuroblastoma cells gave rise to established metastatic sublines. Non-vascularised, vascularised and haemorrhagic areas from the primary tumour xenografts of both models were subcultured, and preliminary results suggest that the haemorrhagic area is the initial seat of metastatic dissemination (Cappellen & Bénard, Institut Gustave Roussy, France). Preliminary data indicate that human neuroblastoma cells are able to metastasise in the nude mouse via the lymphatic route as well as the haematogenous route. These murine models of metastasis originating from human cells may be adequately mimicking human metastatic disease and, thus, be a useful means of studying the genetics of tumour progression and metastases in human neuroblastoma.

Most neuroblastoma lines established *in vitro*, including IGR-N-835 and 91 lines, show *MYCN* amplification. Given the pivotal role played by the *MYCN* oncogene in the expression of the neuroblastoma malignant phenotype, this oncoprotein may be a determinant, not only for oncogenesis, but also for metastasis. There is evidence to support this assumption. Firstly, *in vitro* transfection of stable *MYCN* to SH-EP neuroblasts not expressing *MYCN*, generated advanced malignancy in human neuroblastoma cells, characterised by the expression of an autocrine basic FGF (fibroblast growth factor) loop and angiogenesis [31, 32]. Secondly, IGR-N-91 neuroblasts, cultured *in vitro* from blood cells, bone marrow and the myocardium of mice bearing a subcutaneous tumour IGR-N-91 xenograft, elicited, with consistent *MYCN* amplification, a significant increase in *MYCN* gene transcript levels [30].

GENETIC ALTERATIONS SPECIFICALLY RELATED TO A LACK OF RESPONSE TO CHEMOTHERAPY

If metastatic cells are the essential targets of chemotherapy, then a lack of response to such treatment could signify that the metastatic cancer cell has acquired additional genetic alterations leading to the chemoresistance phenotype. Among the genes possibly activated in chemoresistant cells is the PGY1 (previously termed the MDR1) gene, which encodes the 170 kDa P-glycoprotein (P-gp) [33], a plasma membrane energydependent multidrug efflux "pump", which expels many hydrophobic chemotherapeutic agents from the target cancer cell (anthracyclines, vinca alkaloids). Neuroblastoma is a chemosensitive neoplasm which can become refractory to many drugs able to select or to induce a multidrug resistant phenotype in many patients and, as such, is no longer chemocurable [34]. This is consistent with the frequent overexpression of the PGY1 gene in this tumour, and the fact that approximately 40% of neuroblastoma derive from the medulla adrenal which overexpresses the PGY1 gene at a high level. The clinical significance of PGY1 gene activation has been extensively investigated [35, 36]. In our laboratory, the prognostic value of PGY1 gene expression in neuroblastoma was assessed on a series of 84 patients, taking into account the main known clinical and biological factors of the disease. In the multivariate analysis, only MYCN amplification and PGY1 overexpression remained significantly associated with an increased risk of death. In agreement with a previous report [37], our data strongly suggested that PGY1 gene overexpression is an independent prognostic factor in neuroblastoma [38]. In this series, a subset of very aggressive metastatic neuroblastomas with MYCN amplification, which were refractory to treatment, raised the question of whether PGY1 gene overexpression was due to treatment itself or due to spontaneous tumour progression and metastatic dissemination. These questions were tested experimentally using in vivo models.

We, therefore, decided to study the possible relationship between MYCN and PGY1 gene expression in the nude mouse xenograft model, IGR-N-91, during metastatic dissemination.

All IGR-N-91 cells, either from primary subcutaneous tumour, IGR-N-91, xenografts or metastases, consistently exhibited 60 copies of MYCN per haploid genome. A significant increase in MYCN gene transcript levels was observed in neuroblastoma metastatic cells compared to those of the primary. MYCN overexpression was associated with a significant rise in PGY1 gene mRNA levels leading to a functional P-glycoprotein [30]. This study showed that the MYCN oncogene and PGY1 gene can both be activated during the metastatic process in the absence of any chemotherapy. Increased MYCN expression has also been demonstrated after progression in human neuroblastoma using paired lines derived from patients at diagnosis and during progression [39]. The extra amount of the MYCN transcript present during the migration of metastatic neuroblastoma cells probably activates genes biologically involved in the so-called "super decathlon", a term used to describe the complex pathway used by metastatic cells (traversing the membranes, adhesion, etc).

The PGY1 gene is not the only potential determinant of chemoresistance in neuroblastoma. Resistance to drugs, such as cisplatinum and cyclophosphamide, also used in the treatment of this cancer, can be mediated by increasing the cytoplasmic thiol detoxification pathways via key enzymes, such as glutathione-S-transferases (GST). In our search for GST3 gene activation in advanced neuroblastomas, we found that increased GST3 expression was not an indicator of tumour response to chemotherapy [40]. In contrast, the combined overexpression of GST3 and PGY1 genes appeared to be significantly related to a poor response of the primary tumour [40], a relationship which remains to be demonstrated in metastases. However, given the results obtained with the IGR-N-91 model and patient tumours, pleiotropic activation of various genes responsible for chemoresistance would appear to occur in advanced neuroblastomas. In MYCN activated neuroblastomas, one possibility is that the nuclear oncoprotein, operating as a transcription factor, activates target genes involved in chemoresistance. Consistent with this assumption is our recent finding that the proximal promoter of the PGY1 gene was activated by MYCN in neuroblastoma cell lines (Ferrandis and Bénard, Institut Gustave Roussy, France), thus indicating that an oncogene can trigger PGY1 gene activity.

BASIC AND INTRINSIC MECHANISMS RELATED TO CELL RESPONSE TO CHEMOTHERAPY IN NEUROBLASTOMA

A possible option in a cell's resistance to treatment is its refusal to undergo apoptosis. In some cancer models, a mass of evidence indicates that the relative expressions of TP53 tumour suppressor gene, C-MYC protoncogene and BCL-2 products are intrinsic genetic factors which determine whether cancer cells proliferate or undergo apoptosis, according to external signals. TP53-dependent apoptosis has been shown to modulate the cytotoxicity of anticancer drugs [41] and most of these drugs are able to induce apoptosis [42]. Thus, the idea that interaction between the drug and its target per se is the sole determinant of cell sensitivity to cytotoxic drugs merits re-appraisal. Tumour response to drugs and therapeutic indexes of treatments may be contingent on the thresholding of the expression of proteins involved in apoptosis, i.e., p53, bcl-2 and c-myc [43]. Furthermore, it is suggested that the intrinsic killing power of the drugs may not be as critical as their ability to activate self-destruction in appropriate tumours [44]. Although anticancer drugs can activate late events of apoptosis, there are essential differences in signalling pathways between phamacological cell death and the physiological induction of an active suicide programme [45]. Differences may also stem from the nature of the tumour tissue [46]. Thus, genes implicated in apoptosis must be scrupulously analysed, taking into account each tumour type and the corresponding treatment.

What, in fact, do we know about the status of these genes in neuroblastoma? Clearly, TP53 gene mutations are rare in neuroblastoma tumours [47–49]; and yet TP53 overexpression has been measured in cell lines [50]. Many authors refute or are sceptical about TP53 gene alterations in the aetiology and progression of neuroblastoma because of the absence of TP53 mutations. It is likely that TP53 may, however, be inactivated in a different manner in this tumour (nuclear MDM2 p53-inactivating protein [51, 52], or cytoplasmic localisation of the p53 protein, as seen in breast cancer [53]. Very recent results indicate that, in the absence of MDM2 amplification, most undifferentiated neuroblastomas exhibit increased p53 cytoplasmic content, as assessed by immunohistochemistry. This suggests a defect in the transport of p53 from the cytoplasm to the nucleus [54].

Activation of the BCL-2 oncogene in neuroblastoma in culture is mostly found in immature neuroblasts, while differentiated cells do not express the anti-apoptotic oncogene [55]. Whether this differentiation-dependent BCL-2 expression operates in patients' tumours and metastases remains to be established.

The last key gene involved in the ability of neuroblastoma cells to undergo apoptosis is, of course, MYCN. In our IGR-N-91 cell model, MYCN overexpression associated with the chemoresistant phenotype suggests a possible role of this onco-protein in the apoptosis of neuroblastoma.

In brief, the status of the TP53, BCL-2 and MYCN genes needs to be thoroughly investigated in neuroblastoma cell lines derived from tumours and metastasis to evaluate how apoptosis is involved in the response of cancer cells to chemotherapy.

CONCLUSION

Our knowledge about the genetic alterations involved in the metastatic process of neuroblastoma is still limited. Major anomalies, including DNA content, MYCN amplification and 1p deletion(s), seem to occur prior to metastatic dissemination, and the expression of the so-called modulator genes seems to be more specifically involved in this process.

In the MYCN activated neuroblastoma IGR-N-91 experimental model, data strongly suggest that the oncogene drives neuroblastoma cells to metastatic dissemination as well as emergence of chemoresistance phenotype. This finding throws light on and supports previous observations reported in human lung carcinoma [56] and in metastatic metastatic rhabdomyosarcoma [57]: C-JUN, C-FOS and C-MYC, all oncogenic transcriptional nuclear factors, are activated in parallel with the PGY1 gene and the potential for metastasis. A direct link between drug resistance and metastatic dissemination is thus currently sought [58].

Neuroblastoma cells appear to resist chemotherapy via two types of mechanisms, a general one causing cells to resist apoptosis and specific mechanisms of detoxification. A neuroblastoma cell's ability to undergo apoptosis appears to be dependent on the levels of expression of some key genes, particularly MYCN. Importantly, results obtained from in vivo metastatic models [30] suggest that the MYCN product activates not only angiogenesis and the dissemination process [31, 32], but also

chemoresistance via chemoresistant specific genes, i.e., PGY1 and GST3. As the MYCN product plays a crucial role in progression to aggressive neuroblastoma, we are currently testing the hypothesis that the expression of this oncoprotein and MAX associated partner [59] attains a certain threshold level which determines proliferation, apoptosis, and/or tumour progression and, perhaps also, resistance to treatment.

- Brodeur G, Azar G, Brother M, et al. Neuroblastoma. Effect of genetic factors on prognosis and treatment. Cancer 1992, 70, 1686-1694.
- Delattre O, Michon J, Zucker J, Thomas G. Acquired genetic alterations in neuroblastoma. Clinical implications. Cancer J 1991, 4, 228-235.
- Bourhis J, DeVathaire F, Wilson GD, et al. Combined analysis of DNA ploidy index and N-myc genomic content in neuroblastoma. Cancer Res 1991, 51, 33-36.
- Bourhis J, Dominici C, McDowell H, et al. N-myc genomic content and DNA ploidy in stage IV-S neuroblastoma. J Clin Oncol 1991, 9, 1371-1375.
- Hart I, Easty D. Tumor cell progression and differentiation in metastasis. Semin Cancer Biol 1991, 2, 87-95.
- Fidler I. Critical factors in the biology of human cancer metastasis. Cancer Res 1990, 50, 6130-6138.
- Farber E, Cameron R. Sequential analysis of cancer development. Adv Cancer Res 1980, 31, 125-226.
- Klein G, Klein E. Evolution of tumours and the impact of molecular oncology. Nature 1985, 315, 190–195.
- Weith A, Martinsson T, Cziepluch C, et al. Neuroblastoma consensus deletion maps to 1p36.1-2. Genes Chrom Cancer 1989, 1, 159-166.
- Takeda O, Homma C, Maseki N, et al. There may be two suppressor genes on chromosome 1p closely associated with biologically distinct subtypes of neuroblastoma. Genes Chrom Cancer 1991, 10, 30-39.
- Schwab M, Alitalo K, Klempnauer KB, et al. Amplified DNA with limited homology to myc cellular oncogene is shared by human neuroblastoma cell lines and a neuroblastoma tumour. Nature 305, 245-248
- 12. Seeger R, Brodeur GF, Sather D, et al. Association of multiple copies of the N-myc oncogene with rapid progression of neuroblastomas. New Engl J Med 1985, 313, 1111-1116.
- Nakagawara A, Arima-Nakagawara M, Scavarda NJ, Azar CG, Cantor AB, Brodeur GM. Association between high levels of expression of the TRK gene and favorable outcome in human neuroblastoma. New Engl J Med 1993, 328, 847-854.
- Caron H, Van Sluis P, Van Hoeve M, et al. Allelic loss of chromosome 1p36 in neuroblastoma is of preferential maternal origin and correlates with N-myc amplication. Nature Genet 1993, 4, 187-190.
- Bernards R. N-myc disrupts protein kinase C-mediated signal transduction in neuroblastoma. EMBO J 1991, 10, 1119–1125.
- 16. Kogner P, Barbany G, Björk O, et al. trk mRNA and low affinity nerve growth factor receptor mRNA expression and triploid DNA content in favorable neuroblastoma tumors. In Evans A, Biedler J, Brodeur G, D'Angio G, Nakagawara A, eds. Advances in Neuroblastoma Research 4. New York, Wiley-Liss, 1994, 137-146.
- Tanaka T. Ha-ras p21 in neuroblastoma: a new marker in patient outcome. In Evans A, Biedler J, Brodeur G, D'Angio G, Nakagawara A, eds. Advances in Neuroblastoma Research 4. New York, Wiley-Liss, 1994, 275-280.
- Tanabe KK, Ellis LM, Saya H. Expression of CD44R1 adhesion molecule in colon carcinomas and metastases. *Lancet* 1993, 341, 725-726.
- Shtilvelman E, Bishop J-M. Expression of CD44 is repressed in neuroblastoma cells. Mol Cell Biol 1991, 11, 5446-5453.
- Favrot M, Combaret V, Lasset C. CD44—a new prognostic marker for neuroblastoma. N Engl J Med 1993, 329, 1965.
- Gross N, Beretta C, Peruisseau G, Jackson D, Simmons D, Beck D. CD44H expression by neuroblastoma cells: relation to MYCN amplification and lineage differentiation. Cancer Res 1994, 54, 4238-4242.
- 22. Hailat N, Keim D, Melhem R, et al. High levels of p19/nm23 protein in neuroblastoma are associated with advanced stage disease and with N-myc amplification. J Clin Invest 1991, 88, 341-345.

J. Bénard

 Leone A, Seeger R, Hong C-M, et al. Evidence for nm23 RNA overexpression, DNA amplification and mutation in aggressive childhood neuroblastomas. Oncogene 1993, 8, 855-865.

- Chang C, Zhu X, Thoravai D, et al. nm23-H1 mutation in neuroblastoma. Nature 1994, 370, 335-336.
- Bernards R, Dessain S, Weinberg R. N-myc amplification causes down-modulation in MHC class I antigen expression in neuroblastoma. Cell 1986, 47, 667-674.
- Gross N, Miescher G, Beck D, Favre S, Meyer M. Influence of exogenous MYCN expression on neuroblastoma growth, morphology, antigenic profile and sensitivity to NK- and Lak-effector cells. Proc Am Assoc Cancer Res 1992, 33, A304.
- 27. Versteeg R, Chen C, Schmidt A, et al. Regulation of N-myc and class I HLA in neuroblastoma: tumor suppressor genes involved? In Schwab M, Tonini G-P, Bénard J, eds. Human Neuroblastoma Recent Advances in Clinical and Genetic Analysis. Harwood Academic Publishers, 1992, 128-34.
- Gazitt Y, He Y, Chang L, Koza S, Fisk D, Graham-Pole J. Expression of N-myc, c-myc, and PGY1 proteins in newly established neuroblastoma cell lines: a study by immunofluorescence staining and flow cytometry. Cancer Res 1992, 52, 2957-2965.
- Bettan-Renaud L, Bayle C, Teyssier JR, Bénard J. Stability of phenotypic and genotypic traits during the establishment of a human neuroblastoma cell line, IGR-N-835. *Int J Cancer* 1989, 44, 460-64.
- Ferrandis E, Da Silva J, Riou G, Bénard J. Coactivation of the PGY1 and MYCN genes in human neuroblastoma cells during the metastatic process in the nude mouse. Cancer Res 1994, 54, 2256-2261.
- Schweigerer L, Breit S, Wenzel A, Tsunamoto K, Ludwig R, Schwab M. Augmented MYCN expression advances the malignant phenotype of human neuroblastoma cells: evidence for induction of autocrine growth activity. Cancer Res 1990, 50, 4411-4416.
- Schweigerer L, Fotsis T. Human neuroblastoma cells with enhanced N-myc expression: growth factors and growth inhibitors. In Schwab M, Tonini GP, Bénard J, eds. Human Neuroblastoma: Recent Advances in Clinical and Genetic Analysis. Harwood Academic Publishers, 1993, 135-143.
- Gottesman M, Pastan I. Biochemistry of multidrug resistance mediated by the multidrug transporter. A Rev Biochem 1993, 62, 285-427
- Hartmann O, Pinkerton C, Philip T, Zucker J-M, Breatnach F. Very high-dose cisplatinum and etoposide in children with untreated advanced neuroblastoma. J Clin Oncol 1988, 6, 44-50.
- Bourhis J, Bénard J, Hartmann O, Boccon-Gibod L, Lemerle J, Riou G. Correlation of PGY1 gene expression with chemotherapy in neuroblastomas. J Natl Cancer Inst 1989, 81, 1401-1405.
- Goldstein LJ, Fojo AT, Ueda K, et al. Expression of the multidrug resistance, PGY1, gene in neuroblastomas. J Clin Oncol 1990, 8, 128-136
- Chan H, Haddad G, Thorner P, et al. P-glycoprotein expression as a predictor of the outcome of therapy for neuroblastoma. New Engl J Med 1990, 325, 1608-1614.
- Bénard J, Bourhis J, de Vathaire F, et al. Prognostic value of PGY1
 gene expression in neuroblastoma: results of a multivariate analysis.
 In Evans A, Biedler J, Brodeur G, D'Angio G, Nakagawara A, eds.
 Advances in Neuroblastoma Research 4. New York, Wiley-Liss,
 1994, 111-116.
- Rosen N, Reynolds P, Thiele CJ, Biedler JL, Israel M. Increased N-myc expression following progressive growth of human neuroblastoma. Cancer Res 1986, 46, 4139-4142.
- Bourhis J, Hartmann O, DeVathaire F, et al. Expression of PGY1 and GSTπ genes in 35 advanced neuroblastomas. In Evans A, Biedler J, D'Angio G, Knudson A, Seeger R, eds. Advances in Neuroblastoma Research 3. New York, Wiley-Liss, 1991, 127-134.

- Lowe S, Ruley HE, Jakes T, Housman D. p53-dependent apoptosis modulates the cytotoxicity of anticancer agents. Cell 1993, 74, 957-967.
- Williams G, Smith C. Molecular regulation of apoptosis: genetic controls on cell death. Cell 1993, 74, 777-779.
- Hickman JA. Apoptosis induced by anticancer drugs. Cancer Metastasis Rev 1992, 11, 121–139.
- Fisher DE. Apoptosis in cancer therapy: crossing the threshold. Cell 1994, 78, 539-542.
- Smets L. Programmed cell death (apoptosis) and response to anticancer drugs. Anti Cancer Drugs 1994, 5, 3-9.
- Slichenmyer W, Nelson W, Slebos R, Kastan M. Loss of a p53associated G1 checkpoint does not decrease cell survival following DNA damage. Cancer Res 1993, 53, 4164-4168.
- Imamura J, Bartram CR, Berthold F, Harms D, Nakamura H, Koeffler HP. Mutation of the p53 gene in neuroblastoma and its relationship with N-myc amplification. Cancer Res 1993, 53, 4053-4058.
- 48. Komuro H, Hayashi Y, Kawamura M, et al. Mutations of the p53 gene are involved in Ewing's sarcoma but not in neuroblastoma. Cancer Res 1993, 53, 5284-5288.
- Vogan K, Bernstein M, Leclerc JM, et al. Absence of p53 gene mutations in primary neuroblastoma. Cancer Res 1993, 53, 5269-5276.
- Davidoff AM, Pence JC, Shorter NA, Iglehart JD, Marks JR. Expression of p53 in human neuroblastoma and neuroepithelioma derived cell lines. Oncogene 1992, 7, 127-133.
- Momand J, Zambetti GP, Olson DC, George D, Levine AJ. The mdm-2 oncogene product forms a complex with the p53 protein and inhibits p53-mediated transactivation. Cell 1992, 69, 1237-1241.
- Oliner JD, Kinzler KW, Meltzer PS, George DL, Vogelstein B. Amplification of a gene encoding a p53-associated protein in human sarcomas. *Nature* 1992, 358, 80-83.
- Moll U, Riou G, Levine A. Two distinct mechanisms alter p53 in breast cancer: mutation and nuclear exclusion. *Proc Natl Acad Sci* 1992, 89, 7262-7266.
- Moll U, La Quaglia M, Bénard J, Riou G. Wild-type p53 protein undergoes nuclear exclusion in undifferentiated neuroblastomas but not in differentiated tumors. Proc Natl Acad Sci USA (in press).
- Hanada M, Krajewski S, Tanaka S, et al. Regulation of Bcl-2 oncoprotein levels with differentiation of human neuroblastoma cells. Cancer Res 1993, 53, 4978

 –4986.
- Volm M. P-glycoprotein associated expression of c-fos and c-jun products in human lung carcinomas. Anticancer Res 13, 375-378.
- Hanania N, Boyano M-D, Mangin C, Poupon M-F. Oncogene and PGY1 gene expression in rat rhabdomyosarcoma sublines of different metastatic potential. Anticancer Res 11, 473

 –480.
- Kerbel RS, Kobahyashi H, Graham CH. Intrinsic or acquired drug resistance and metastasis: are they linked phenotypes? J Cell Biochem 56, 37-47.
- Wenzel A, Cziepluch C, Hamann U, Schürmann J, Schwab M. The N-myc oncoprotein is associated in vivo with the phosphoprotein Max(p20/22) in human neuroblastoma cells. EMBO 3 1991, 10, 3703-3712

Acknowledgements—I thank Marie-France Poupon, Institut Curie, Paris for critical reading of the manuscript and Lorna Saint-Ange, Institut Gustave Roussy for editing the manuscript. The work performed at the Institut Gustave Roussy and partly reported in this review was supported by Association de la Recherche sur le Cancer, Villejuif and CRC Institut Gustave Roussy No. 94-3, Villejuif. I thank collaborators Jaqueline Da Silva, Eric Ferrandis, Nathalie Duarte and David Cappellen for their contribution, Guy Riou (Molecular Pharmacology Laboratory, Head), Oliver Hartmann (Bone Marrow Transplantation Unit, Chief) and Jean Lemerle (Paediatric Department, Head), Institut Gustave Roussy-Villejuif for interest and support of this work.